Competitive seedlings and inherited traits: a test of rapid evolution of *Elymus multisetus* (big squirreltail) in response to cheatgrass invasion

Courtney L. J. Rowe and Elizabeth A. Leger

1 USDA Forest Service, Plumas National Forest, Quincy, CA, USA
2 Department of Natural Resources and Environmental Science, University of Nevada, Reno, NV, USA

Introduction

The impact of invasive species on native species, communities, and ecosystem processes has been recognized for decades (Elton 1958; Lodge 1993). Biologic invasions dramatically affect the distribution, abundance, and fitness of many native species (Williamson 1996; Mack et al. 2000). While biologic invasions are considered a significant threat to biodiversity (Sala et al. 2000; Stein et al. 2000), they do not result in the extinction of all native species impacted; many native species are able to persist alongside invaders (Hironaka and Tisdale 1963; Levine and Rees 2004; Mealor et al. 2004; Stohlgren et al. 2006). These persistent native species may possess traits (i.e. preadaptations) or sufficient phenotypic plasticity that allows for successful coexistence with invaders (MacNeil et al. 2001; Carroll et al. 2005). Preadapted traits can increase in frequency in invaded populations, or under the right circumstances, native species may evolve new mechanisms, such as chemical defenses or improved competitive ability, to cope with the invaders. Both types of rapid evolutionary responses have been documented in native populations (Schlaepfer et al. 2005; Strauss et al. 2006). Most research involving rapid evolution in native species responding to invaders has focused on native–invader interactions across trophic levels (e.g. predator–prey, host–pathogen relationships, Strauss et al. 2006). To date, there are fewer studies that address evolutionary response of native plant populations to plant invaders (e.g. Callaway et al. 2005; Lau 2006; Mealor and Hild 2007; Cipollini and Hurley 2008; Leger 2008; Ferrero-Serrano et al. 2010), but all have found evidence that...
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some native plants can evolve in response to invasive competitors. The experiment presented here differs from previous ones in that, in addition to determining whether native populations have evolved increased ability to compete with invaders, we also identify particular traits that are evolving, as well as determine the amount of genetic variation within populations for adaptive morphological and phenological traits.

The conversion of Great Basin rangeland to Bromus tectorum L. (cheatgrass) is one of the most severe ecological degradations in the United States (Mack 1981; D’Antonio and Vitousek 1992). A combination of heavy grazing, which reduces the vigor of native perennial species, and B. tectorum invasion, which shortens fire return interval and alters nutrient cycling, has resulted in large-scale conversion of Artemisia spp. (sagebrush)-dominated plant communities to B. tectorum-dominated communities (Young et al. 1972; Evans et al. 2001; Brooks et al. 2004; Chambers et al. 2007). By 2000, nearly 12.7 million hectares of the Great Basin had been converted to B. tectorum and 45% was estimated to be at risk of conversion (Menakis et al. 2003; Bradley and Mustard 2005).

Bromus tectorum has strong negative impacts on native perennial plant fitness (Aguirre and Johnson 1991; Nasri and Doescher 1995; Rafferty and Young 2002; Humphrey and Schupp 2004). A few native grass species appear to be relatively tolerant of B. tectorum invasion, including the closely related E. multisetus M.E. Jones (big squirreltail) and Elymus elymoides Raf. (bottlebrush squirreltail) (Hironaka and Tisdale 1963; Booth et al. 2003; Leger 2008). Both are highly selfing, short-lived perennials that reproduce effectively from seed, persist in disturbed areas, and survive moderate fires, with gene flow occurring primarily through wind-dispersed seeds (Young and Evans 1977; Britton et al. 1990; Jones 1998; Larson et al. 2003). A tolerance for disturbance has made both squirreltail species of interest for the restoration of burned and B. tectorum-invaded areas (Jones 1998; Richards et al. 1998; Humphrey and Schupp 2002). Direct comparison between squirreltail and annual grass competitors in relative growth rate (Hironaka and Sindelar 1975; Arrendondo et al. 1998), productivity under both low and high N availability (James 2008), and germination percentage and phenology (Young et al. 2003) usually favor the invader, suggesting that no individual strategy can account for squirreltail’s tolerance of invasion.

Increased ability to tolerate B. tectorum competition has been demonstrated in mature E. multisetus plants collected from B. tectorum-invaded areas (Leger 2008), but shifts in seedling performance have not been investigated. The transition from seed to seedling is the most vulnerable stage of plant life (Kitajima 2007) and is crucial for a successful restoration from seed; examination of seedling performance is an important measure for estimating fitness differences between plants. Any trait with inherited phenotypic variation that increases seedling establishment in B. tectorum-invaded areas would likely be favored by natural selection and increase in frequency over time in invaded populations. Measuring the performance of seeds from the same plant (i.e. siblings) can provide an estimate of genetic variation in particular traits. Family-level analysis has been used for decades to address the questions of inheritance and natural selection (Endler 1986; Falconer and MacKay 1996) but is not frequently applied in the context of native plant restoration.

The goals of this study are twofold: (i) to test whether seedlings of E. multisetus plants collected from invaded areas are more successful when grown with B. tectorum than seedlings from nearby uninvaded areas and (ii) to identify what growth traits may help E. multisetus seedlings to establish in B. tectorum-invaded areas. We conducted a common garden greenhouse experiment with maternal families collected from invaded and uninvaded areas with known differences in adult competitive ability (Leger 2008), and we measured E. multisetus seedling performance and B. tectorum biomass production. Two methodologies were used to identify potentially important traits. First, we identified traits that were correlated with increased tolerance of B. tectorum competition in seedlings, quantified genetic variation of these traits, and asked whether adaptive traits have shifted between uninvaded and invaded populations. If populations are evolving in response to B. tectorum pressure and if traits correlated with performance in the greenhouse are also important in the field, we would expect to see an increased frequency of inherited performance-related traits within invaded areas. Secondly, we surveyed all growth traits that varied in frequency between uninvaded and invaded populations, casting a wider net to identify additional traits that may confer a competitive advantage to native plants growing in invaded areas.

Materials and methods

Seed collection methods

Seeds of E. multisetus were collected in June 2007 from a sagebrush steppe plant community in Balls Canyon, Sierra County, California (Fig. 1.; 139840.740 N, 120803.360 W, 1600 m elevation), where a shift in invasion status occurs over a relatively small geographic area (ranging from 0% B. tectorum in uninvaded areas to 40% B. tectorum cover in invaded areas). Detailed description of community composition of the collection area is recorded in Leger (2008). At the time of seed collection, B. tectorum invasion at this site followed fence line boundaries, with B. tectorum occurring in an area that has received a
longer annual season of higher intensity grazing than uninvaded areas for ~100 years (C. Ross, personal communication). To ensure that the greatest differences between collections were disturbance history and current *B. tectorum* cover, rather than other abiotic conditions, all collection sites were located within 1.2 km of each other (Fig. 1). However, there are uncontrolled environmental differences between invaded and uninvaded collection sites, namely elevation and soil type. At Balls Canyon, invaded seed collection sites are, on average, 36 m lower than uninvaded areas. Higher elevation sites also exhibit different soil types than lower elevation sites; while all series are well drained and exhibit the same bedrock, the soils at higher elevation collection sites contain a higher percentage of coarse material (>2 mm) (NRCS 2008). A portion of the study area burned in 2006, and the remainder burned in 2007, and currently, the density and cover of *B. tectorum* in uninvaded areas sampled for this study has increased (E. Leger, personal observation), indicating that the lack of invasion in the higher elevation sites was probably due to differences in disturbance history, rather than lack of suitable environmental conditions for invasion.

Seeds were collected from individual *E. multisetus* plants from four invaded and uninvaded sites (Fig. 1) and stored at room temperature. All seeds collected from an individual represent a maternal family line, and a ‘family’ refers to seeds collected from an individual plant in the field. Twenty-five families were randomly selected from the available pool of seeds from invaded and uninvaded sites (50 families total). Depending on seed availability, 18–20 seeds from each family were used for this experiment (991 plants total). For brevity, families and seedlings produced from seeds collected in invaded sites are subsequently referred to as ‘invaded families’ and ‘invaded plants’ and families and seedlings produced from seeds collected at uninvaded sites, ‘uninvaded families’ and ‘uninvaded plants.’ Seeds of *B. tectorum* were bulk-collected from the invaded portion of the study area in June 2007.

**Greenhouse and data collection methods**

The experiment was conducted under controlled greenhouse conditions: 4.4–26.6°C temperature range; 5–25% relative humidity; full daytime sunlight. Potting mix was locally produced (R.C. Donovan, Reno, NV) and contained bark, compost, decomposed granite, and perlite. Laboratory tests of the potting mix revealed very low mineral N (3 ppm), high alkalinity (pH = 8.0), and low estimated water holding capacity (44%) (A and L Western Agricultural Laboratories Report #08-136-060 2008). To allow for maximum rooting depth, we used pots designed for revegetation horticulture (Stuewe and Sons TFOT1; 10.2 × 10.2 × 35.6 cm; 3.2 L). Seeds received no pregermination treatment, and one individually weighed *E. multisetus* seed was directly sown into each pot on 26 March 2008. To create competitive conditions, we sowed five *B. tectorum* seeds into each of 14–16 pots for each squirreltail family (*n* = 791). This density was chosen to produce a competitive environment but was lower than field densities to facilitate root harvests (described elsewhere). An additional four pots per family were established as noncompeting controls (*n* = 200). Pots for the same family were sown together and then moved into randomly assigned positions to achieve a complete random design. We misted the soil surface immediately after sowing and continued misting twice daily for 2 weeks. For the remainder of the experiment, plants were hand-watered to saturation when the bottom third of root zone was dry (4- to 14-day intervals), which maintained relatively droughty conditions, and no supplemental fertilizer was added.

We measured or calculated a total of 47 growth traits including above-ground and below-ground measures (root methods are described in detail below): seed weight; emergence date; early growth rate (leaf number and length); root, shoot, and total biomass weight (mg) at 10, 50, and 100 days; leaf number and root to shoot ratio.
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(R:S) at 10, 50, and 100 days; total root length (cm), root diameter (mm), number of root tips, number of root forks, and root length in five 0.1 mm size classes at 10, 50, and 100 days after peak germination. Three sequential harvests were conducted on seedlings grown in competition with B. tectorum. Harvest dates were selected to correspond to important phenological benchmarks, including seedling establishment phase (10 days after peak germination), the active growth phase (50 days), and the end of growing season (100 days). Plants for harvest were randomly selected within families. Two plants per family (n = 100) were harvested at 10 days after peak germination, (14–16 April 2008), and three plants per family (n = 150) were harvested at 50 days (16–18 May 2008). At 100 days, owing to the time requirements for root harvest, we were only able to harvest one competing and one noncompeting plant per family (n = 100, 15–18 July 2008). At 100 days, leaf number was recorded for all live plants (n = 669). Plant traits were deliberately measured at a common time, rather than at a particular age/size class, to measure E. multisetus performance relative to B. tectorum in a realistic seedling competition scenario (B. tectorum seeds typically emerge within 24–36 h of planting, which was the case in this experiment as well). Thus, differences between plants harvested on the same date may reflect patterns determined by ontology of trait development (e.g. Fransen et al. 1999; Aanderud et al. 2003) as well as fixed differences in traits.

Emergence began on 1 April 2008 and was recorded daily until last recorded emergence on 15 April 2008. By 4 April 2008, 820 of 921 seeds had emerged, which represented over 90% of all emergents, and all seedling ages are displayed from 4 April, representing the end of peak germination. For a subset of 406 seedlings, leaf length and number were measured on 23 April and 15 May 2008. Early seedling growth rate was quantified as [May leaf number–April leaf number]/May leaf number. To extract roots at harvest, we cut away pots and gently removed planting media by misting with water. Elymus multisetus were identified by leaf and root characteristics and manually separated from B. tectorum roots. Harvested roots and shoots were refrigerated (<48 h) and then digitally scanned for analysis using WinRhizo software (Regent Instruments Inc, Saint-Foy, QC). WinRhizo was utilized to quantify the following root characteristics: total root length (cm), root diameter (mm), number of root tips and root forks (measurements of root branching), and root length (cm) in five 0.1-mm-diameter size classes. Root and shoot biomass measurements were taken by drying (7 days at 60°C) and weighing root and shoot biomass separately. Total biomass was calculated as shoot + root weight. No plants produced seeds under competition in the second year of growth, so common-garden-produced seed was collected and weighed from a subset of noncompeting control plants from each family that were maintained through July 2009.

Relative competitive performance index (CPI) was used to quantify seedling tolerance to B. tectorum competitors (Keddy et al. 1998). CPI is the percent decrease in plant performance when grown with competitors and was calculated as: [Leaf number without competition–leaf number with competition]/leaf number without competition. Leaf number is strongly correlated with plant biomass in E. multisetus (presented in results) and was used as a proxy for plant size in CPI calculation. We calculated family means for leaf number with and without competition, and these mean values were used for CPI calculation. To quantify B. tectorum suppression, above-ground biomass was collected from all pots containing B. tectorum (n = 507) at 100 days. In 32 competition pots, there was no E. multisetus emergent, but four or more B. tectorum emergents. Elymus multisetus seeds that failed to emerge were significantly smaller than seeds that did emerge (presented in results), indicating that failure of E. multisetus emergence in some pots was likely related to seed factors, rather than to soil characteristics or pot placement. Pots without E. multisetus seedlings were used to calculate mean B. tectorum weight grown alone, and B. tectorum shoots were dried and weighed in same manner as E. multisetus.

Analysis methods

Statistical analyses were performed using JMP 7.0.2 (SAS Institute, Cary, NC, USA). Analysis of variance (ANOVA) was used to compare the effects of invasion status and family on 47 growth traits of E. multisetus seedlings in competition. For seed weight, emergence date, early growth rates, 10-day harvest data, 50-day harvest data, 100-day leaf number, and next-generation seed weight, the ANOVA model included the following factors: invasion status (invaded or uninvaded); collection site (random factor, nested in status); family (random factor, nested in status and site). Family was removed from ANOVA models for the analysis of 100-day biomass and 100-day root traits, because only one plant per family was measured in each competition condition, as well as from ANOVA of CPI, because they were calculated using family means. Seed weight and emergence date comparisons included all data, rather than just competing plants, because these traits were not affected by competition condition. For B. tectorum biomass, two separate ANOVAs were conducted to analyze the effect of three levels of competition condition (grown alone, or grown with
invaded/uninvaded *E. multisetus* and the family of *E. multisetus* competitor. For this second analysis of *B. tectorum* biomass, *E. multisetus* family was nested within collection site. When analyses were significant, means were compared *a posteriori* using Tukey Honestly significant difference (HSD). Several growth traits were log-transformed to improve fit to assumptions of ANOVA (indicated in Table 1). Raw means and standard errors, rather than transformed values or least means square, are reported for all growth trait and competitive ability measurements. Spearman’s \( \rho \) was used to determine nonparametric correlations between family mean CPI and family means for each growth trait among uninvaded families. No adjustments were made for multiple comparisons in these analyses because the goal of these experiments was to maximize the chances of identifying potentially important traits, which can serve as the foundation for additional follow-up testing in this and other native perennial grass species.

### Results

At 100 days, *E. multisetus* plants grown in competition with *B. tectorum* had 81.4% less total biomass (\( P < 0.0001, F_{1,100} = 46.88, r^2 = 0.32 \)), 70% less root biomass (\( P < 0.0001, F_{1,100} = 33.15, r^2 = 0.25 \)), 90.2% less shoot biomass (\( P < 0.0001, F_{1,101} = 51.80, r^2 = 0.34 \)), and 82.5% fewer leaves (\( P < 0.0001, F_{1,672} = 708.12, r^2 = 0.51 \)) than control plants grown without competition. At 100 days, leaf number was highly correlated with total biomass (\( P < 0.0001, F_{1,100} = 10005.48, r^2 = 0.91 \)), root biomass (\( P < 0.0001, F_{1,100} = 223.42, r^2 = 0.70 \)), and shoot biomass (\( P < 0.0001, F_{1,101} = 10005.48, r^2 = 0.91 \)). Fifty-day leaf number was also highly

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**Table 1.** Traits of *Elymus multisetus* grown in competition with *Bromus tectorum* that varied significantly by invasion status, family, or both.

<table>
<thead>
<tr>
<th>Response variable</th>
<th>Status</th>
<th>Site</th>
<th>Family</th>
</tr>
</thead>
<tbody>
<tr>
<td>Seed weight (g)</td>
<td>991</td>
<td>0.44</td>
<td>69.77</td>
</tr>
<tr>
<td>Emergence date</td>
<td>950</td>
<td>0.09</td>
<td>0.08</td>
</tr>
<tr>
<td>Early growth rate (leaf no.)</td>
<td>402</td>
<td>0.47</td>
<td>33.93</td>
</tr>
<tr>
<td>Early growth rate (leaf length)</td>
<td>402</td>
<td>0.29</td>
<td>11.91</td>
</tr>
<tr>
<td>10-day traits</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>10-day total biomass (mg)</td>
<td>99</td>
<td>0.62</td>
<td>0.01</td>
</tr>
<tr>
<td>10-day shoot biomass (mg)</td>
<td>147</td>
<td>0.61</td>
<td>7.09</td>
</tr>
<tr>
<td>10-day root biomass (mg)</td>
<td>148</td>
<td>0.66</td>
<td>5.22</td>
</tr>
<tr>
<td>10-day R:S</td>
<td>147</td>
<td>0.50</td>
<td>6.40</td>
</tr>
<tr>
<td>10-day leaf no.</td>
<td>148</td>
<td>0.66</td>
<td>8.42</td>
</tr>
<tr>
<td>10-day root total length (cm)*</td>
<td>145</td>
<td>0.53</td>
<td>0.11</td>
</tr>
<tr>
<td>10-day root tips*</td>
<td>145</td>
<td>0.50</td>
<td>2.69</td>
</tr>
<tr>
<td>10-day root forks*</td>
<td>145</td>
<td>0.56</td>
<td>1.50</td>
</tr>
<tr>
<td>10-day root diameter (mm)*</td>
<td>145</td>
<td>0.64</td>
<td>58.21</td>
</tr>
<tr>
<td>10-day % fine roots (&lt;0.2 mm)</td>
<td>145</td>
<td>0.65</td>
<td>40.05</td>
</tr>
<tr>
<td>10-day (&gt;0.1 mm)*</td>
<td>145</td>
<td>0.57</td>
<td>0.12</td>
</tr>
<tr>
<td>10-day (0.101–0.2 mm)*</td>
<td>145</td>
<td>0.49</td>
<td>3.91</td>
</tr>
<tr>
<td>10-day (0.301–0.4 mm)*</td>
<td>145</td>
<td>0.53</td>
<td>3.67</td>
</tr>
<tr>
<td>10-day (&gt;0.4 mm)*</td>
<td>145</td>
<td>0.64</td>
<td>37.90</td>
</tr>
<tr>
<td>100-day traits</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>100-day total biomass (mg)</td>
<td>49</td>
<td>0.46</td>
<td>7.45</td>
</tr>
<tr>
<td>100-day shoot biomass (mg)</td>
<td>50</td>
<td>0.42</td>
<td>7.22</td>
</tr>
<tr>
<td>100-day root biomass (mg)</td>
<td>49</td>
<td>0.44</td>
<td>6.39</td>
</tr>
<tr>
<td>100-day leaf no.</td>
<td>482</td>
<td>0.57</td>
<td>66.34</td>
</tr>
</tbody>
</table>

Sites represent collection locations within invaded and uninvaded areas. At 100 days, root, shoot, and total weights were only measured for one plant per family, and therefore, family is not indicated as an analysis factor for these traits; this is indicated by ‘–’. Numerator degrees of freedom for the factors status, site, and family are 1, 6, and 42, respectively. Significant factors (\( \alpha = 0.05 \)) are highlighted in bold. Asterisks demark traits whose values were log-transformed for analysis.

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correlated with 50-day total root, and shoot biomass (all $P < 0.001$). However, at 10 days, leaf number was not significantly correlated with any biomass measure (all $P > 0.05$), as most plants had ≤3 leaves at this time, regardless of their size.

**CPI and Bromus tectorum suppression**

Competitive performance index varied significantly by invasion status ($P = 0.0160$, $F_{1,50} = 6.30$, $r^2 = 0.31$) and collection site ($P = 0.0496$, $F_{6,50} = 2.33$, $r^2 = 0.31$). Mean CPI of invaded families was significantly lower than that of uninvaded families (Fig. 2), indicating that plants from invaded areas (4.78 ± 0.039 mg, Table 1), and these invaded plants still produced smaller seeds (5.2 ± 0.04 mg) than uninvaded plants (6.1 ± 0.04 mg). Additionally, at each harvest date, invaded plants exhibited lower root, shoot, and total biomass, with significant differences in shoot and total biomass at 50 days and in all three measures at 100 days (Table 1; Fig. 5A–C). Invaded plants exhibited significantly lower early leaf growth rates (0.33 ± 0.013 leaves added; 0.26 ± 0.013 cm leaf length gained, Table 1) than uninvaded plants (0.45 ± 0.018 leaves added; 0.39 ± 0.035 cm leaf length gained, Table 1). Invaded plants exhibited higher R:S at 50 and 100 days, but not at 10 days; this difference was significant at 50 days (Table 1; Fig. 5D). At both 50 and 100 days, invaded plants had significantly lower leaf numbers than uninvaded plants (Tables 1 and 2). The average root diameter of invaded plants at 50 days was significantly smaller than that of uninvaded plants (Tables 1 and 2).

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**Figure 2** Relative competitive performance index (CPI) of Elymus multisetus plants that differ in their exposure to Bromus tectorum in the field. Values are means ± SE. Letters represent significant differences between plants from invaded and uninvaded areas ($\alpha = 0.05$). Plants with lower CPI values are better tolerators of competition, showing a smaller reduction in biomass when grown with B. tectorum.

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**Figure 3** Bromus tectorum shoot biomass production when grown with Elymus multisetus seedlings that differ in invasion history. Values are means ± SE, and letters represent significant differences between groups ($\alpha = 0.05$). Lower B. tectorum weight signifies a greater suppressive effect and is seen in plants collected from invaded areas.

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**Growth traits in a competitive environment**

When grown in competition with B. tectorum, E. multisetus seedlings exhibited 14 growth traits that varied significantly by invasion status and 19 growth traits that varied significantly by family (Table 1). Of the 23 growth traits that varied significantly by invasion status or family, 61 (14 traits) were found in the 50-day harvest group when we had increased statistical power because of greater sample size, while none were from the 10-day harvest group. Field-collected seeds from invaded areas exhibited significantly lower seed weights (4.32 ± 0.046 mg) than uninvaded plants (4.78 ± 0.039 mg, Table 1), and these differences persisted in seeds collected from noncompeting control plants grown in common garden ($P < 0.0001$, $F_{1,200} = 209.72$, $r^2 = 0.91$). Seeds produced in the common garden were larger than field-collected seeds, but invaded plants still produced smaller seeds (5.2 ± 0.04 mg) than uninvaded plants (6.1 ± 0.04 mg). Additionally, at each harvest date, invaded plants exhibited lower root, shoot, and total biomass, with significant differences in shoot and total biomass at 50 days and in all three measures at 100 days (Table 1; Fig. 5A–C). Invaded plants exhibited significantly lower early leaf growth rates (0.33 ± 0.013 leaves added; 0.26 ± 0.013 cm leaf length gained, Table 1) than uninvaded plants (0.45 ± 0.018 leaves added; 0.39 ± 0.035 cm leaf length gained, Table 1). Invaded plants exhibited higher R:S at 50 and 100 days, but not at 10 days; this difference was significant at 50 days (Table 1; Fig. 5D). At both 50 and 100 days, invaded plants had significantly lower leaf numbers than uninvaded plants (Tables 1 and 2). The average root diameter of invaded plants at 50 days was significantly smaller than that of uninvaded plants (Tables 1 and 2).

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**Table 1** Competitive performance index (CPI) and biomass of Bromus tectorum seedlings grown in competition with Elymus multisetus. Values are means ± SE, and letters represent significant differences between groups ($\alpha = 0.05$). Lower B. tectorum weight signifies a greater suppressive effect and is seen in plants collected from invaded areas.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>CPI</th>
<th>Shoot biomass (g)</th>
<th>Root biomass (g)</th>
<th>Total biomass (g)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Grown alone</td>
<td>75%</td>
<td>1.50 ± 0.02</td>
<td>0.50 ± 0.01</td>
<td>2.00 ± 0.03</td>
</tr>
<tr>
<td>Grown with invaded</td>
<td>74%</td>
<td>1.25 ± 0.02</td>
<td>0.35 ± 0.01</td>
<td>1.60 ± 0.03</td>
</tr>
<tr>
<td>Grown with uninvaded</td>
<td>73%</td>
<td>1.75 ± 0.02</td>
<td>0.65 ± 0.01</td>
<td>2.40 ± 0.03</td>
</tr>
</tbody>
</table>

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**Table 2** Growth traits of Bromus tectorum seedlings grown in competition with Elymus multisetus. Values are means ± SE, and letters represent significant differences between groups ($\alpha = 0.05$).

<table>
<thead>
<tr>
<th>Trait</th>
<th>Growth</th>
<th>Invaded</th>
<th>Uninvaded</th>
<th>$P$-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Leaf number</td>
<td>50 days</td>
<td>3.0 ± 0.5</td>
<td>4.0 ± 0.5</td>
<td>$&lt; 0.0001$</td>
</tr>
<tr>
<td>Root diameter</td>
<td>100 days</td>
<td>0.5 ± 0.1</td>
<td>0.6 ± 0.1</td>
<td>$&lt; 0.0001$</td>
</tr>
<tr>
<td>Biomass</td>
<td>100 days</td>
<td>2.0 ± 0.1</td>
<td>2.5 ± 0.1</td>
<td>$&lt; 0.0001$</td>
</tr>
</tbody>
</table>

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and 2). The total length of large diameter roots (>0.4 mm) varied significantly by invasion status, with uninvaded plants exhibiting greater length of large diameter roots than invaded plants (Tables 1 and 2). Finally, at each harvest date, invaded plants exhibited higher percentages of fine roots than uninvaded plants, with significant differences at 50 days (Tables 1 and 2). Four root diameter size classes exhibited detectable family-level differences at 50 days, while only one family-level difference in root distribution was detected at 10 days (Table 1).

**Traits correlated with CPI**

Within uninvaded families, three growth traits were significantly correlated with CPI: 10-day R:S ($P = 0.0178$),
10-day root forks (\(P = 0.0427\)), and 10 day length of small diameter roots (0.101–0.200 mm, \(P = 0.0309\)).

Lower CPI (i.e. greater competitive ability) was correlated with higher 10-day R:S, more root forks, and greater total length of small diameter roots (Fig. 6). While 10-day root forks did not vary significantly between families when considering plants from both invaded and uninvaded areas (Table 1), among only uninvaded plants, 10-day root forks varied significantly by family (\(P = 0.0224\), \(F_{21,49} = 2.13\), \(r^2 = 0.70\)). Neither 10-day R:S nor the length of small diameter roots varied between families within uninvaded plants. Analysis of all harvested plants did not detect differences between invaded and uninvaded plants for 10-day root forks, R:S, or small diameter (0.101–0.200 mm) roots. For these three traits, power analysis revealed large sample size requirements (>1000) to detect invasion status differences.

**Influence of seed weight on performance**

Several early growth traits were significantly affected by seed weight. Larger seeds were more likely to emerge (\(P < 0.0001\), \(F_{1,989} = 30.20\), \(r^2 = 0.03\); nonemergent: 3.8 ± 0.1 mg; emergent: 4.6 ± 0.02 mg). Ten-day root biomass (\(P < 0.0001\), \(F_{2,99} = 28.13\), \(r^2 = 0.22\)), shoot biomass (\(P < 0.0001\), \(F_{2,100} = 26.62\), \(r^2 = 0.22\)), and total biomass (\(P < 0.0001\), \(F_{2,99} = 36.10\), \(r^2 = 0.27\)) increased with increasing seed weight. The length of roots in the three largest diameter classes root classes also increased with increasing seed weight (all \(P < 0.02\)). No 50-day or 100-day growth traits were affected by seed weights (all \(P > 0.05\)). Neither CPI nor *B. tectorum* biomass were significantly by family, indicating genetic variation for this trait exists in natural populations. Although the difference in *B. tectorum* tolerance was small-only a 3% decrease in CPI for invaded plants, *B. tectorum* suppression was 30% higher for invaded plants than uninvaded plants. We do not know whether these differences are sufficient to confirm the hypothesis that *B. tectorum* is exerting a strong selection pressure on native grass populations in the Great Basin: we found that *E. multisetus* seedlings from invaded areas were more tolerant of competition and better able to suppress *B. tectorum* than plants from nearby uninvaded areas. In the Great Basin, *B. tectorum* invasion is associated with other disturbances such as heavy grazing and fire. Therefore, trait differences observed across the invasion boundary in this study may result from additional factors that affect plant fitness, including grazing history or soil type, both of which are known to differ between invaded and uninvaded collection sites. Nonetheless, as confirmed by our results, competition with *B. tectorum* is known to have strong effects on the fitness of perennial native plants; wholesale invasion certainly has the ability to impose strong selection on such plants. Regardless of the number of selective agents involved, results from this type of experiment can identify plants that perform well in the presence of *B. tectorum* and that may offer the best potential for restoring invaded rangelands.

The ability to suppress *B. tectorum* biomass varied significantly by family, indicating genetic variation for this trait exists in natural populations. Although the difference in *B. tectorum* tolerance was small-only a 3% decrease in CPI for invaded plants, *B. tectorum* suppression was 30% higher for invaded plants than uninvaded plants. We do not know whether these differences are sufficient to

**Table 2.** Values for growth traits of *Elymus multisetus* grown in competition with *Bromus tectorum* by invasion status from three different harvest dates.

<table>
<thead>
<tr>
<th>Trait</th>
<th>10 day Uninvaded</th>
<th>10 day Invaded</th>
<th>50 day Uninvaded</th>
<th>50 day Invaded</th>
<th>100 day Uninvaded</th>
<th>100 day Invaded</th>
</tr>
</thead>
<tbody>
<tr>
<td>Leaf number</td>
<td>1.22 (0.06)</td>
<td>1.20 (0.06)</td>
<td>8.04 (0.60)</td>
<td>6.32 (0.46)</td>
<td>10.84 (0.36)</td>
<td>7.81 (0.27)</td>
</tr>
<tr>
<td>Root length (cm)</td>
<td>17.12 (0.97)</td>
<td>16.80 (0.87)</td>
<td>671.18 (44.97)</td>
<td>684.01 (43.54)</td>
<td>1067.57 (107.53)</td>
<td>1160.37 (154.01)</td>
</tr>
<tr>
<td>Root tips</td>
<td>49.92 (5.12)</td>
<td>48.80 (3.88)</td>
<td>1535.03 (130.21)</td>
<td>1389.32 (120.84)</td>
<td>4026.08 (467.44)</td>
<td>3717.69 (538.23)</td>
</tr>
<tr>
<td>Root forks</td>
<td>49.22 (3.78)</td>
<td>51.88 (4.87)</td>
<td>3361.81 (364.69)</td>
<td>3926.24 (365.90)</td>
<td>7143.76 (1022.56)</td>
<td>8096.50 (1537.62)</td>
</tr>
<tr>
<td>Root diameter (mm)</td>
<td>0.32 (0.01)</td>
<td>0.31 (0.01)</td>
<td>0.26 (0.01)</td>
<td>0.21 (0.01)</td>
<td>0.23 (0.01)</td>
<td>0.21 (0.01)</td>
</tr>
<tr>
<td>% fine roots (&lt;0.2 mm)</td>
<td>0.34 (0.02)</td>
<td>0.39 (0.02)</td>
<td>0.72 (0.01)</td>
<td>0.78 (0.01)</td>
<td>0.74 (0.01)</td>
<td>0.78 (0.01)</td>
</tr>
<tr>
<td>Root diameter size classes (root length in cm)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>&lt;0.1 mm</td>
<td>1.86 (0.22)</td>
<td>1.68 (0.15)</td>
<td>216.22 (20.91)</td>
<td>237.86 (21.85)</td>
<td>423.41 (49.67)</td>
<td>451.23 (66.27)</td>
</tr>
<tr>
<td>0.101–0.2 mm</td>
<td>4.09 (0.37)</td>
<td>4.92 (0.37)</td>
<td>269.78 (15.24)</td>
<td>295.20 (14.42)</td>
<td>359.96 (28.81)</td>
<td>451.00 (52.80)</td>
</tr>
<tr>
<td>0.201–0.3 mm</td>
<td>2.60 (0.19)</td>
<td>2.49 (0.20)</td>
<td>46.24 (3.19)</td>
<td>49.78 (3.15)</td>
<td>81.22 (8.42)</td>
<td>78.60 (10.14)</td>
</tr>
<tr>
<td>0.301–0.4 mm</td>
<td>6.37 (0.43)</td>
<td>5.65 (0.37)</td>
<td>46.56 (3.62)</td>
<td>49.34 (2.94)</td>
<td>80.49 (8.79)</td>
<td>80.12 (9.86)</td>
</tr>
<tr>
<td>&gt;0.4 mm</td>
<td>2.15 (0.21)</td>
<td>2.03 (0.24)</td>
<td>90.67 (8.86)</td>
<td>50.29 (5.62)</td>
<td>119.38 (19.02)</td>
<td>96.32 (19.74)</td>
</tr>
</tbody>
</table>

Values are untransformed means (SE). Significant differences between traits are displayed in Table 1.
provide a measurable advantage to *E. multisetus* from invaded populations when growing in a natural setting. Field testing is underway to determine whether the lower CPI and greater ability to suppress *B. tectorum* exhibited by invaded seedlings will result in greater survival of these plants in invaded areas.

The findings from both our correlative analysis and our examination of shifts in traits suggest that root growth plays a significant role in *E. multisetus* seedling performance when in competition with *B. tectorum*. Our most conservative approach for finding adaptive traits was a three-step process of assessing which traits were correlated with CPI within the presumed ancestral gene pool (i.e. uninvaded areas), determining the level of genetic variation in these traits, and examining whether trait values have shifted in the invaded populations. Of the 47 growth traits measured, three were correlated with CPI and all were related to root growth at 10 days. Although type I statistical errors are possible when analyses are conducted on a large number of traits, the fact that all CPI-correlated traits were identified from the earliest seedling stage suggests that these relationships are biologically important, rather than statistical artifacts. Greater tolerance to competition was associated with greater fine root production, higher R:S, and more root forking in young seedlings (Fig. 6). We did not detect a shift across invasion status for these traits, indicating either that traits have not shifted in response to *B. tectorum* or that we lacked the power to detect a shift. Identifying traits that are easier to measure and are reasonable proxies for root traits (e.g. resource capture) could increase sample sizes and improve the resolution of this method. Differences in root traits were also observed in our broader comparative analysis. Invaded plants produced a significantly higher percentage of fine roots and roots of smaller average diameter at 50 days (Tables 1 and 2). At 50 and 100 days, invaded plants allocated a relatively greater proportion of biomass to root production (Fig. 5).

When competition is mostly for soil resources (e.g. in situations with low soil fertility or high water stress), allocation to below-ground resource capture (measured by high R:S, high specific root length, high fine root production) improves plant performance (McGrath and Chapin 1989; Aerts et al. 1991; Tilman and Wedin 1991; Casper and Jackson 1997; Aerts and Chapin 2000). Several mechanisms have been suggested to explain the importance of fine roots in resource acquisition, including improved opportunistic nutrient foraging (Fitter 1994), adventitious root production in ephemeral nutrient patches (Drew et al. 1973; Robinson et al. 1999), and enhanced nitrogen uptake at terminal roots (Pregitzer et al. 2002). Fine root production and increased root biomass have been linked to resource capture and competitive ability in Great Basin plants (e.g. Bilbrough and Caldwell 1997; James et al. 2009). Root forks may contribute to increased competitive ability in nutrient-poor environments, where root morphology rather than physiology may confer the greatest advantage in nutrient uptake ability (Aerts 1999).
For example, in an examination of genetic variation of root morphology in white clover, it was suggested that genotypes with a high number of root tips and forks will possess highly branched root systems that can explore a larger volume of soil per unit root weight, thereby improving nutrient uptake (Jahufer et al. 2008).

Both alone and in competition with B. tectorum, invaded plants were smaller than uninvaded plants. They possessed fewer leaves (Tables 1 and 2), had lower total, root, and shoot biomass (Table 1; Fig. 5), and added biomass more slowly. Nonetheless, seedlings from plants collected from B. tectorum-invaded areas were better at suppressing B. tectorum. This result is contrary to the prevailing theory that greater plant size confers greater competitive ability (Gaudet and Keddy 1988; Keddy et al. 2002). There are other plant–plant interactions besides direct competition that can result in the appearance of competitive advantage. For example, it is possible that some plants reduce the size of their neighbors releasing an allelopathic compound or stimulating the growth of soil organisms that inhibit plant growth, rather than superior resource capture (Allen and Allen 1986; Mahall and Callaway 1992). Although we did not investigate these mechanisms, they may account for our observed disassociation between biomass and competitive ability.

Beyond seeing an unexpected negative relationship between E. multisetus size and competitive ability, we observed that B. tectorum produced significantly more biomass when grown with E. multisetus from uninvaded areas than when it was grown alone, potentially indicative of a facilitative effect (Fig. 3). Interactions between plants are increasingly recognized to contain facilitative components through mechanisms such as soil aeration, root exudation (which can facilitate nutrient availability or alter biotic soil communities), or amelioration of abiotic stress (reviewed in Brooker et al. 2008). We do not know which, if any, of these mechanisms are responsible for the apparent facilitative effect of larger E. multisetus seedlings from uninvaded areas on B. tectorum, but a similar facilitative relationship has been observed in interactions between other desert annual forbs and their competitors (E. Leger, unpublished data). Results of this experiment suggest that a mechanism more complex than a positive relationship between size and competitive ability may be involved in interactions between plants in arid systems.

Our finding that smaller native plants possessed greater tolerance to B. tectorum competition than larger plants may not be that unusual; there can be adaptive value in being a small plant (Aarssen et al. 2006), especially in desert systems. Size does not always predict the impact of resource limitation: in a study of six annual species, greater biomass in isolation did not confer advantage in offspring production under competition; in fact, the most fecund species under competition exhibited relatively small biomass in isolation (Neytcheva and Aarssen 2008). Through its high seed production and soil water exploitation, B. tectorum can drastically reduce resources for native grass seedling establishment (Melgoza et al. 1990), creating highly competitive conditions where small size may be advantageous. Furthermore, in arid systems where plants are often water stressed, there can be disadvantages to greater biomass (e.g. higher transpiration and tissue maintenance costs) that result in mortality; smaller seedlings have shown a greater ability to survive drought conditions (Hendrix et al. 1991).

Invaded plants were not only smaller seedlings; they also had smaller seeds than uninvaded plants. Within a species, bigger seeds typically produce bigger seedlings, though the effect often weakens after development of the first photosynthetic tissue (e.g. Stanton 1984; Kitajima 2007). Our results were consistent with this pattern, with larger seeds producing larger seedlings for both invaded and uninvaded sources at 10 days, while initial seed weight was not significantly related to B. tectorum tolerance, B. tectorum suppression, or any 50- or 100-day growth traits, indicating that the importance of seed weight diminished over time. Nongenetic maternal provisioning can influence seed size (Roach and Wulff 1987); however, it is unlikely that larger seed size of uninvaded plants was due purely to maternal effects, as seed size differences persisted in seed collected from plants grown in a common garden, which is consistent with some degree of genetic control. Seed size is not the only growth trait that can be influenced by maternal environment, and as plants evaluated in this experiment were field collected, there are other maternal environment effects that may have affected performance which were not evaluated here. Given the large variation in family performance (Table 1, Fig. 4), even among plants that were experiencing the same maternal environment, it is likely that there was a genetic component to the differences we found between families and populations, and between invaded and uninvaded areas. We observed family-level variation at a high frequency, which is evidence that many growth traits are inherited (Table 1). Genetic variation in fitness-related traits is the basis for natural selection, and greater genetic variation represents increased evolutionary potential and a greater likelihood of long-term population persistence (Frankel 1974; Stockwell et al. 2003). Elymus multisetus and E. elymoides are known to have a high degree of morphological variation among populations (Jones et al. 2003), relatively high amplified fragment length polymorphism (AFLP) diversity within populations compared to other perennial grass species ( Larson et al. 2003), and our work demonstrates that E. multisetus also has a large
amount of family-level genetic variation in growth traits. Because these grasses occur in a wide range of habitats across the Great Basin and are relatively tolerant of disturbance (Jones 1998), it is possible that the large amount of variation observed within these species is a result of their relatively large range sizes, persistence under recent disturbance, and high population densities. Populations with low genetic diversity may not possess sufficient evolutionary potential to persist under new sources of disturbance, such as climate change, land use changes, or new biologic invasions (Rice and Emery 2003; Harris et al. 2006). Although not all the inherited traits we measured were related to performance in greenhouse conditions, the level of genetic variation that we detected may improve the chances for long-term persistence of E. multisetus populations in a changing environment.

The ability of native populations to evolve in response to B. tectorum invasion has significant implications for the management and restoration of invaded communities. If long-invaded populations possess increased tolerance and competitive ability with B. tectorum (Figs 2 and 3), then seed from invaded populations could be an effective tool in restoring nearby B. tectorum-degraded areas. However, seed selection need not be limited to long-invaded populations, as there exists significant genetic variation in how seedlings respond to B. tectorum within uninvaded populations (Fig. 4). Screening plants for inherited performance-related traits and the subsequent choice of genotypes or populations that exhibit high tolerance or suppression of B. tectorum for use in restoration may improve restoration success. Additional traits may be important for survival, growth, and reproduction in B. tectorum-invaded areas, including germination timing, fire and/or grazing tolerance, and disease resistance, which also warrant investigation.

One final consideration for land managers is that restoration projects often introduce large volumes of seed into an ecosystem and may affect both the quantity and quality of local genetic variation. Commercially available seed is often derived from a single source population and sometimes even from a single genotype (Young et al. 2003; Jones and Larson 2005; Shaw et al. 2005). Because nonlocal genotypes can have a deleterious effect on the fitness of local populations (Waser et al. 2000; Montalvo and Ellstrand 2001), there is widespread concern about genetic swamping during large-scale restoration (Montalvo et al. 1997; Hufford and Mazer 2003; McKay et al. 2005). The impact of large-scale seeding on the genetic integrity of existing native populations and the ecosystem as a whole remains poorly understood (Knapp and Dyer 1998; Broadhurst et al. 2008). If B. tectorum-invaded populations are rapidly evolving novel genotypes, then the introduction of high quantities of nonlocal seed near long-invaded populations may reduce both their fitness and evolutionary potential.

Conclusions

The invasion of B. tectorum is converting large areas of the Great Basin from native perennial-dominated to B. tectorum-dominated plant communities; however, some native plants are able to persist in these invaded areas and may evolve in response to long-term B. tectorum presence. Our findings were consistent with the hypothesis that B. tectorum is exerting strong selective pressure on native grass populations in the Great Basin. This study provides examples of two methods for assessing potentially adaptive growth traits: (i) directly testing which traits correlate with increased performance, in either greenhouse or field settings; (ii) comparing traits of successful populations (identified by persistence in invaded areas) with plants growing in uninvaded areas. Both methods have the potential to identify important traits for persistence of native plants in invaded, disturbed, or altered environments. The ability to identify which traits are beneficial and the degree of genetic variation in these traits will help to advance our understanding of the mechanisms of contemporary evolutionary change. It can also serve a practical function by informing management and restoration of native populations in invaded or otherwise disturbed areas.

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References


